

Synthetic studies of thiazoline and thiazolidinecontaining natural products – 2. Total synthesis of the antimycoplasma antibiotic micacocidin

Akira Ino,* Yasushi Hasegawa, and Akira Murabayashi

Aburahi Laboratories, Shionogi & Co., Ltd., Koka, Shiga 520-3423, Japan

Received 15 April 1999; accepted 25 June 1999

Abstract

Synthesis of the right half of micacocodin (segment B) and subsequent completion of total synthesis of the antimycoplasma antibiotic micacocidin is described. The desired S-configuration at C-14 secondary carbinol was obtained by stereoselective reduction of the preceding ketone in accordance with the Cram rule. Condensation of two labile segments, A and B, was achieved in the presence of potassium acetate. The chiral center at C-10 was finally isomerized to the natural configuration through formation of the Zn complex. © 1999 Elsevier Science Ltd. All rights reserved.

Keywords: antibiotics; thiazolines; thiazolidines; chelation.

In our preceding paper [1], we reported synthesis of protected segment A (2) of the antimycoplasma antibiotic micacocidin (1) [2], in which phosphorus pentachloride-mediated thiazoline constructing reaction was applied. In this paper, we describe the synthesis of the right half of 1 (segment B), and condensation of these two segments leading to 1 [3].

Scheme 1. Retrosynthetic analysis

To introduce three chiral centers into segment B, we planned to use cysteine for C-12 and C-18, while stereoselective reaction was used for C-14 configuration. Construction of the central thiazolidine ring through condensation of segments A and B, and stereochemical control on the configuration at the C-10 carbon were performed with reference to model experiments described in the previous report [1]. Thus, we retrosynthesized segment B to L-cysteine (3a), isobutyric acid (4) and 2-methyl-S-cysteine (5) (Scheme 1). An initial attempt starting with N-methyl-L-cysteine (3b) was unsuccessful because the N-methyl-oxazolidinone ring corresponding to 14 resisted cleavage [4], so herein we began the synthetic study with 3a.

Synthesis of segment B

Elongation of the carboxyl moiety in thiazolidine 6 [5], which was prepared from L-cysteine (3a) via a two-step reaction, was achieved by carbonyldiimidazole treatment and subsequent condensation with methyl isobutyrate to yield keto-ester 7. Reduction of 7 with sodium borohydride proceeded with sufficient stereoselectivity in accordance with the Cram rule as expected to yield the desired S-alcohol 8, along with a trace amount of R isomer and an over-reduced diol 11, both of which were readily separated by chromatography. Compound 8 was then converted to 9 through construction of an oxazolidinone ring with sodium hydride and subsequent hydrolysis. The configuration of 9 was confirmed by NOE examination as shown in Scheme 2. When the elongation reaction was performed with aldehyde 10 [6] and methyl isobutyrate, the unwanted R-alcohol was predominant (R:S=ca.3:2 by NMR).

$$\begin{array}{c} \text{Me} \quad \text{S} \\ \text{Me} \quad \text{N} \\ \text{Boc} \\ \text{CO}_2\text{H} \\ \end{array} \begin{array}{c} 1. \; (\text{Imid.})_2\text{CO} \\ \hline 2. \; \text{Me}_2\text{CHCO}_2\text{Me} \\ \hline 75\% \\ \end{array} \begin{array}{c} \text{Me} \quad \text{N} \\ \text{Boc} \\ \hline 7 \\ \end{array} \begin{array}{c} \text{Me} \quad \text{NaBH}_4 \\ \hline \text{Me} \quad \text{Me} \quad \text{Me} \\ \hline \text{S}_{9\%} \\ \end{array} \begin{array}{c} \text{Me} \quad \text{Me} \quad \text{Me} \\ \hline \text{S}_{9\%} \\ \hline \text{Me} \quad \text{Me} \\ \hline \text{S}_{9\%} \\ \end{array} \begin{array}{c} \text{Me} \quad \text{Me} \quad \text{Me} \\ \hline \text{S}_{9\%} \\ \hline \text{Me} \quad \text{Me} \quad \text{Me} \\ \hline \text{Me} \quad \text{Me} \\ \hline \text{Me} \quad \text{Me} \quad \text{Me} \quad \text{Me} \\ \hline \text{Me} \quad \text{Me} \quad \text{Me} \quad \text{Me} \\ \hline \text{Me} \quad \text{Me} \quad$$

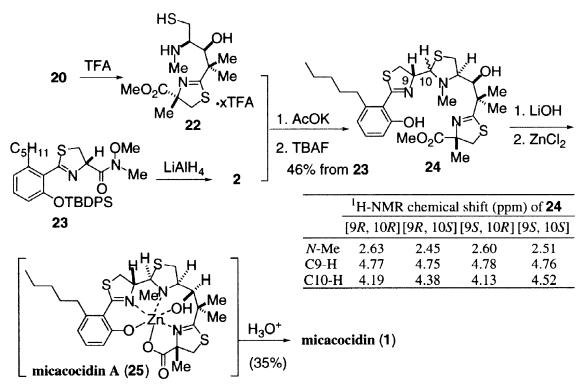
Next, condensation of 9 with 2-methyl-S-cysteine methyl ester hydrochloride [7] was performed using carbonyldiimidazole to yield peptide 12. Treatment of 12 with trifluoroacetic acid in refluxing toluene accomplished cyclization of the N-acylcysteine moiety with concomitant removal of the acetonide protecting moiety. Since the methyl ester moiety was also hydrolyzed during the reaction, the product was treated with (trimethylsilyl)diazomethane to yield thiol 13. When the cyclization reaction of 12 was carried out under dehydration conditions, for example with a Dean-Stark trap, the major product was 21, and attempts to remove the acetonide moiety resulted in simultaneous cleavage of the thiazoline ring. Then, the thiol residue in 13 was protected with a p-methoxybenzyl (PMB) group.

Cleavage of the oxazolidinone ring in 14 was performed as follows. Introduction of a t-butoxycarbonyl (Boc) group and subsequent treatment with methanolic cesium carbonate [8], followed by remethylation of the hydrolyzed methoxycarbonyl group, afforded 15 with concomitant recovery of 14. After protection of newly formed secondary alcohol with a t-butyldimethylsilyl (TBS) group, N-methylation of 16 was effected by treatment with

dimethyl sulfate and sodium hydride. Since recovered 16 was not readily separable from 17 by chromatography, the product contaminated with 16 was then treated with a small amount of tetrabutylammonium fluoride (TBAF) at low temperature to selectively desilylate 16. Pure 17 thus obtained was afresh treated with TBAF in the presence of 4A molecular sieves (4A-MS), followed by remethylation of the hydrolyzed ester to afford 18. Regeneration of the thiol residue in 18 was achieved by substituting a 3-nitro-2-pyridinesulfenyl (Npys) group for PMB and subsequent treatment with tributylphosphine [9] to provide Bocprotected segment B 20. (Scheme 3)

Total synthesis of micacocidin (1)

The Boc group in 20 was removed by treatment with trifluoroacetic acid to give amino-thiol 22 which was ready for condensation reaction. On the other hand, as mentioned in our preceding report [1], the TBDPS-protected segment A (2) was fairly labile, so that segment A was preserved as Weinreb amide 23. Condensation of alkaline sensitive 22 and acid sensitive 2, generated from 23 by LiAlH₄ treatment immediately before use, was achieved in the presence of potassium acetate, and subsequent desilylation with TBAF afforded micacocidin methyl ester 24 as a mixture of diastereomers involving C-10 (main) and C-9 configurations.



Scheme 4.

After hydrolysis of the terminal ester group with LiOH, treatment of the resulting acid with zinc chloride resulted in isomerization of the C-10 configuration intensifying natural chirality via micacocidin A (25) formation $(9R:9S = \text{ca.3:1}, \text{ by } ^{1}\text{H-NMR})$. Finally, purification of the product by HPLC furnished micacocidin (1). (Scheme 4)

Synthetic micacocidin thus obtained was identified with the natural sample by comparison of their behavior on HPLC and spectroscopic properties (IR, NMR, MS, $[\alpha]_D$).

Experimental section

General. H-NMR spectra were recorded on a JEOL GSX-270 or JMN A-400 spectrometer, and other instruments used to obtain physical data and experimental conditions for chromatography were the same as described in the preceding paper [1].

Ketone 7. To an ice-cold solution of acid 6 [5] (16.53 g) in THF (62.0 ml) was added carbonyldiimidazole (10.0 g, 1.00 eq.). The mixture was stirred at the same temperature for 30 min and then at room temperature for 2 h. The reaction mixture was diluted with AcOEt, washed with water and brine. The AcOEt layer was dried over Na_2SO_4 and concentrated under reduced pressure to give crude acylimidazole (18.15 g).

Under a nitrogen atmosphere, methyl isobutyrate (14.0 ml, 2.10 eq.) was added dropwise over 10 min to a solution prepared by adding LDA (43.7 ml of 2.0 M, 1.50 eq.) at -78°C into Et₂O (140 ml). This mixture was stirred at the same temperature for 25 min. Then, the crude acylimidazole (18.15 g) solution in THF (100 ml) was added over 30 min, and this mixture was stirred at the same temperature for a further 30 min. The reaction was quenched with 10 % aq.citric acid, and the mixture was extracted with AcOEt. The organic layer was taken and washed with water and brine, dried over Na₂SO₄, then concentrated under reduced pressure. The residue was purified by chromatography (silica 500 g, AcOEt/hexane = 1/6) to give 7 (16.45 g, 75%). Mp 45-48 °C; $[\alpha]^{24}_{D}$ -59.2 (c 1.01, CHCl₃); IR v_{max} 2976, 2933, 1717, 1683, 1363, 1171 cm⁻¹; ¹H-NMR (270 MHz, CDCl₃) δ 1.44 (3H, s), 1.45 (9H, s), 1.54 (3H, s), 1.75 (3H, br-s), 1.83 (3H, br-s), 2.92 (1H, br-s), 3.23 (1H, br-s), 3.74 (3H, s), 5.29 (1H, br-s); Anal. calcd for $C_{16}H_{27}NO_{5}S$: C, 55.63; H, 7.88; N, 4.05. found: C, 55.48; H, 7.77; N,4.18.

Secondary alcohol 8. To an ice-cold solution of ketone **7** (16.45 g, 47.6 mmol) in EtOH (71.0 ml) was slowly added NaBH₄ (1.80 g, 1.00 eq.), and this mixture was stirred at room temperature overnight. The reaction mixture was poured into ice-cold sat.aq.NH₄Cl slowly, and extracted with AcOEt. The organic layer was washed with brine, dried over Na₂SO₄, then concentrated under reduced pressure. The residue was chromatographed (silica 500 g, AcOEt/hexane = 1/6 to 1/2) to afford **8** (9.80 g, 59%) and over-reduced diol **11** (3.69 g, 24%). **8**; Mp 60-63 °C; $[\alpha]_{D}^{24}$ -40.0 (c 1.00, CHCl₃); IR ν_{max} 3483, 2977, 2933, 1727, 1694,

1657, 1366, 1170 cm⁻¹; ¹H-NMR (270 MHz, CDCl₃) δ 1.25 (3H, s), 1.31 (3H, s), 1.48 (9H, s), 1.78 (6H, s), 2.38 (1H, d, J= 12.2 Hz), 3.17 (1H, dd, J= 12.2, 5.5 Hz), 3.38 (1H, br-s), 3.70 (3H, s), 3.90 (1H, t, J= 9.2 Hz), 4.56 (1H, dd, J= 9.2, 5.5 Hz); Anal. calcd for $C_{16}H_{29}NO_5S$: C, 55.31; H, 8.41; N, 4.03. found: C, 55.11; H, 8.03; N,4.21. 11; Mp 128-129 °C; $[\alpha]_{D}^{24}$ -16.8 (c 1.00, CHCl₃); IR v_{max} 3367, 3254, 2971, 2868, 1662, 1394, 1168 cm⁻¹; ¹H-NMR (270 MHz, CDCl₃) δ 0.98 (3H, s), 1.06 (3H, s), 1.51 (9H, s), 1.80 (3H, s), 1.81 (3H, s), 2.48 (1H, d, J= 12.0 Hz), 3.12 (1H, br-s), 3.29 (1H, dd, J= 12.0, 5.3 Hz), 3.47 (1H, t, J= 8.1 Hz), 3.56 (1H, dd, J= 11.7, 5.8 Hz), 3.64 (1H, dd, J= 11.7, 7.4 Hz), 3.86 (1H, br-s), 4.74 (1H, dd, J= 8.1, 5.3 Hz); EIMS m/z 639 [2M+H]⁺, 320 [M+H]⁺, 264, 220; HR-EIMS m/z 320.1909 [M+H]⁺ (calcd 320.1896 for $C_{15}H_{30}NO_4S$); Anal. calcd for $C_{15}H_{29}NO_4S$: C, 56.40; H, 9.15; N, 4.38. found: C, 56.19; H, 9.00; N,4.34.

Carboxylic acid 9. To an ice-cold solution of alcohol 8 (9.80 g, 28.2 mmol) in THF (70.0 ml) was slowly added NaH (60%, 1.35 g, 1.20 eq.), and the mixture was stirred at room temperature for 1.5 h. The reaction mixture was poured into ice-cold sat.aq.NH₄Cl slowly, and the whole was extracted with AcOEt. The organic layer was washed with brine, dried over Na₂SO₄, then concentrated under reduced pressure. The residue was purified by chromatography (silica 320 g, AcOEt/hexane = 1/4) to yield a white solid (methyl ester of 9, 5.54 g).

A mixture of the solid (5.54 g, 20.3 mmol) and NaOH (0.97 g, 1.20 eq.) in MeOH (30.0 ml) and water (10.0 ml) was stirred under reflux overnight. After cooling to room temperature, MeOH was removed in vacuo, and the residue was taken up in water and washed with AcOEt. The aqueous phase was acidified (pH 3-4) with aq.HCl, and then extracted twice with AcOEt. The combined organic phase was washed with brine, dried over Na₂SO₄, then concentrated under reduced pressure to afford pure **9** (4.87 g, 67 % from **8**) as a white solid. Recrystallization of the solid from AcOEt-hexane gave colorless crystals (3.59 g). Mp 182-185 °C; $[\alpha]^{24}_D$ -5.0 (c 1.01, MeOH); IR v_{max} 3273, 2978, 2938, 1735, 1370, 1264 cm⁻¹; ¹H-NMR (270 MHz, CDCl₃) δ 1.30 (3H, s), 1.38 (3H, s), 1.69 (3H, s), 1.98 (3H, s), 3.04 (1H, dd, J = 10.4, 9.8 Hz), 3.12 (1H, dd, J = 10.4, 5.5 Hz), 4.31 (1H, ddd, J = 9.8, 6.1, 5.5 Hz), 4.45 (1H, d, J = 6.1 Hz); Anal. calcd for $C_{11}H_{17}NO_4S$: C, 50.95; C, 50.95; C, 50.95; C, 50.84; C, 50.84; C, 50.84; C, 50.84; C, 50.84; C, 8.31 (18.00) and 18.00 (19.00) and 19.00 (19.00) and 19.00

Peptide 12. An ice-cold solution of acid **9** (3.89 g, 15.0 mmol) in THF (30.0 ml) was treated with carbonyldiimidazole (2.55 g, 1.05 eq.), and stirred at room temperature for 1.5 h. This mixture containing crude acylimidazole was added to an ice-cold solution of 2-Me-S-CysOMe•HCl [7] (4.18 g, 1.5 eq.) in DMF (30.0 ml) over 5 min. After stirring at room temperature for 3 h, the reaction was quenched with sat.aq.NH₄Cl, and extracted with AcOEt. The organic phase was taken and washed with brine, dried over Na₂SO₄, then evaporated under reduced pressure. The residue was purified by chromatography (silica 270 g, AcOEt/hexane = 2/3) to afford **12** (4.40 g, 75%). [α]²⁸_D -40.6 (c 1.00, CHCl₃); IR ν _{max} 3363,

2980, 2934, 2567, 1743, 1657, 1520, 1371, 1260 cm⁻¹; ¹H-NMR (270 MHz, CDCl₃) δ 1.29 (3H, s), 1.32 (3H, s), 1.60 (3H, s), 1.61 (1H, dd, J = 9.2, 5.5 Hz), 1.68 (3H, s), 1.95 (3H, s), 2.99 (1H, dd, J = 11.0, 9.2 Hz), 3.07 (1H, dd, J = 11.0, 5.5 Hz), 3.15 (1H, dd, J = 14.0, 8.6 Hz), 3.35 (1H, dd, J = 14.0, 9.2 Hz), 3.80 (3H, s), 4.36 (2H, m), 6.79 (1H, br-s); LSIMS m/z 781 [2M+H]⁺, 391 [M+H]⁺, 331, 219; HR-LSIMS m/z 391.1360 [M+H]⁺ (calcd 391.1361 for $C_{16}H_{27}N_2O_5S_2$).

Thiol 13. A mixture of 12 (4.00 g, 10.2 mmol), TFA (10.0 ml) and toluene (90.0 ml) was heated with stirring under reflux for 3 days. After cooling to room temperature, the reaction mixture was concentrated under reduced pressure. The residue was dissolved in Et₂O (45.0 ml) and MeOH (5.00 ml), and treated with (trimethylsilyl)diazomethane (2.00 M in hexane) dropwise until generation of N₂ gas ceased. After quenching the methylation with AcOH, the reaction mixture was diluted with AcOEt, washed with sat.aq.NH₄Cl and brine, dried over Na₂SO₄, and then evaporated under reduced pressure. The residue was purified by chromatography (silica 150 g, AcOEt /hexane = 2/1) to afford 13 as an oil (2.22 g, 65%). [α]²⁴_D -2.0 (c 0.50, CHCl₃); IR ν_{max} 3270, 2978, 2934, 2563, 1741, 1606, 1236 cm⁻¹; ¹H-NMR (270 MHz, CDCl₃) δ 1.28 (3H, s), 1.36 (3H, s), 1.49 (1H, dd, J = 10.1, 7.8 Hz), 1.51 (3H, s), 2.58 (1H, ddd, J = 13.9, 10.1, 7.8 Hz), 2.90 (1H, ddd, J = 13.9, 7.8, 3.3 Hz), 3.12 (1H, d, J = 11.4 Hz), 3.66 (1H, d, J = 11.4 Hz), 3.79 (3H, s), 3.80 (1H, m), 4.47 (1H, d, J = 3.8 Hz), 5.77 (1H, br-s); EIMS m/z 333 [M+H]⁺, 201; HR-EIMS m/z 333.0930 [M+H]⁺ (calcd 333.0943 for C₁₃H₂₁N₂O₄S₂).

Thiazolidine 21. A mixture of **12** (391 mg, 1.00 mmol), TFA (1.00 ml) and toluene (9.00 ml) was heated with stirring under reflux for 3 days. During the reaction, H₂O generated *in situ* was removed with an equipped Dean-Stark trap. After cooling to room temperature, the reaction mixture was concentrated under reduced pressure. The residue was dissolved in AcOEt, washed with sat.aq.NaHCO₃ and brine, dried over Na₂SO₄, and then evaporated under reduced pressure. The residue was purified by chromatography (silica 15 g, AcOEt /hexane = 1/2 to 1/1) to afford **21** (207 mg, 56%) and **13** (100 mg, 30 %). **21**; [α]²⁴_D -7.2 (c 0.50, CHCl₃); IR v_{max} 2979, 2936, 1759, 1610, 1370, 1236 cm⁻¹; ¹H-NMR (270 MHz, CDCl₃) δ 1.30 (3H, s), 1.37 (3H, s), 1.53 (3H, s), 1.67 (3H, s), 1.97 (3H, s), 3.00 (1H, dd, J=10.4, 9.8 Hz), 3.10 (1H, dd, J=10.4, 5.5 Hz), 3.12 (1H, d, J=11.6 Hz), 3.68 (1H, d, J=11.6 Hz), 3.78 (3H, s), 4.35 (1H, dt, J=9.8, 5.5 Hz), 4.46 (1H, d, J=5.5 Hz); EIMS m/z 373 [M+H]⁺, 201; HR-EIMS m/z 373.1259 [M+H]⁺ (calcd 373.1256 for $C_{16}H_{25}N_2O_4S_7$).

Thiazoline 14. To an ice-cold solution of **13** (2.22 g, 6.68 mmol) in DMF (13.4 ml) was added p-methoxybenzyl chloride (1.09 ml, 1.20 eq.) and K_2CO_3 (1.38 g, 1.50 eq.), and the mixture was stirred at room temperature for 2 h. The reaction mixture was diluted with AcOEt, washed with water and brine, dried over Na_2SO_4 , and then evaporated under reduced pressure. The residue was chromatographed (silica 140 g, AcOEt /hexane = 3/2) to give **14**

(2.64 g, 87%). $[\alpha]_D^{28}$ -19.8 (c 1.00, CHCl₃); IR ν_{max} 3283, 2971, 2941, 1751, 1609, 1511, 1248 cm⁻¹; ¹H-NMR (270 MHz, CDCl₃) δ 1.23 (3H, s), 1.29 (3H, s), 1.46 (3H, s), 2.49 (1H, dd, J = 13.4, 9.2 Hz), 2.83 (1H, dd, J = 13.4, 3.7 Hz), 3.09 (1H, d, J = 11.6 Hz), 3.64 (1H, d, J = 11.6 Hz), 3.70 (2H, s), 3.74 (1H, m), 3.77 (3H, s), 3.80 (3H, s), 4.36 (1H, d, J = 3.7 Hz), 5.14 (1H, br-s), 6.86 (2H, d, J = 8.6 Hz), 7.22 (2H, d, J = 8.6 Hz); LSIMS m/z 905 [2M+H]⁺, 453 [M+H]⁺, 331, 121; HR-LSIMS m/z 453.1523 [M+H]⁺ (calcd 453.1518 for $C_{21}H_{29}N_2O_5S_2$).

Thiazoline alcohol 15. A mixture of oxazolidinone 14 (2.50 g, 5.52 mmol), (Boc)₂O (1.33 g, 1.10 eq.) and DMAP (34.0 mg, 0.05 eq.) in CH₂Cl₂ (16.6 ml) was stirred at room temperature for 30 min, and then the mixture was concentrated under reduced pressure. A solution of the residue in MeOH (27.6 ml) was treated with Cs₂CO₃ (3.60 g, 2.00 eq) and stirred at room temperature for 22 h. The reaction was quenched with 10% aq.citric acid and extracted twice with AcOEt. The combined organic phase was washed with brine, dried over Na₂SO₄, and then evaporated under reduced pressure. A solution of the residue in Et₂O (45.0 ml) and MeOH (5.00 ml) was treated with ethereal diazomethane, which was prepared from N-methyl-N'-nitro-N-nitrosoguanidine and KOH, dropwise at 0°C, until generation of N₂ gas ceased. The reaction was quenched with AcOH, and concentrated under reduced pressure to yield a residue which was chromatographed (silica 100 g, AcOEt /hexane = 1/4 to 2/1) to afford 15 (878 mg, 30%) and recovered 14 (1.64 g, 66%). 15; $[\alpha]_{D}^{28}$ -29.4 (c 1.00, CHCl₃); IR v_{max} 3418, 3295, 2973, 2932, 1739, 1705, 1609, 1512, 1249, 1173 cm⁻¹; ¹H-NMR (270) MHz, CDCl₃) δ 1.24 (3H, s), 1.36 (3H, s), 1.43 (9H, s), 1.57 (3H, s), 2.52 (1H, dd, J = 14.0, 5.5 Hz), 2.67 (1H, dd, J = 14.0, 8.5 Hz), 3.08 (1H, d, J = 11.6 Hz), 3.61 (1H, d, J = 11.6 Hz), 3.73 (2H, s), 3.78 (3H, s), 3.79 (3H, s), 3.86 (1H, br-s), 3.95 (1H, br-q, J=9.7 Hz), 5.17 (1H, br-qbr-d, J = 10.4 Hz), 5.48 (1H, br-s), 6.83 (2H, d, J = 8.6 Hz), 7.29 (2H, d, J = 8.6 Hz); LSIMS m/z 527 [M+H]⁺, 230, 202, 121; HR-LSIMS m/z 527.2244 [M+H]⁺ (calcd 527.2250 for $C_{25}H_{39}N_2O_6S_2$).

N-Me-TBS-ether 17. To a solution of alcohol **15** (436 mg, 0.83 mmol) and 2,6-lutidine (0.38 ml, 4.00 eq.) in CH_2Cl_2 (4.10 ml) was added TBSOTf (0.38 ml, 2.00 eq.) dropwise over 5 min at -78°C, and the mixture was stirred at the same temperature for 30 min. The reaction was quenched with sat.aq.NaHCO₃ and extracted with AcOEt. The AcOEt extract was washed with water and brine, dried over Na₂SO₄, and then evaporated under reduced pressure. The residue was purified by chromatography (silica 25 g, AcOEt /hexane = 1/5) to afford **16** (479 mg, 90%). IR v_{max} 3453, 2954, 2932, 1735, 1711, 1610, 1512, 1250, 1175 cm⁻¹; ¹H-NMR (270 MHz, CDCl₃) δ 0.10 (3H, s), 0.11 (3H, s), 0.92 (9H, s), 1.23 (3H, s), 1.27 (3H, s), 1.44 (9H, s), 1.56 (3H, s), 2.49 (1H, dd, J = 13.4, 7.9 Hz), 2.60 (1H, dd, J = 13.4, 6.1 Hz), 3.10 (1H, d, J = 11.6 Hz), 3.65 (1H, d, J = 11.6 Hz), 3.71 (2H, d, J = 2.4 Hz), 3.77 (3H, s), 3.78 (3H, s), 3.92 (1H, br-q, J = 7.9 Hz), 4.17 (1H, br-s), 5.09 (1H, br-d, J = 8.5 Hz), 6.82 (2H, d, J = 8.5 Hz), 7.27 (2H, d, J = 8.5 Hz).

To a solution of 16 (479 mg, 0.75 mmol) and Me_2SO_4 (942 mg, 10.0 eq.) in DMF (7.50 ml) was slowly added NaH (60%, 239 mg, 8.00 eq.). The mixture was stirred at room temperature for 30 min and then at 75 °C overnight. After cooling to room temperature, the reaction mixture was diluted with AcOEt and Et₂O, and then the organic phase was washed with sat.aq.NH₄Cl, water and brine, dried over Na₂SO₄, and concentrated under reduced pressure. The residue was dissolved in THF (6.00 ml) and treated with TBAF (1.00 M in THF, 0.19 ml, 0.25 eq.) at 0° C, and stirred at the same temperature for 20 min. The reaction mixture was poured into 5% aq.KHSO₄ and extracted with AcOEt. The AcOEt extract was washed with brine, dried over Na, SO₄, then evaporated under reduced pressure. The residue was purified by chromatography (silica 25 g, AcOEt /hexane = 1/5 to 1/3) to give 17 (304) mg, 62%) and recovered 15 (55.0 mg, 14%). 17; $[\alpha]^{24}$ -26.4 (c 0.50, CHCl₃); IR ν_{max} 2955, 2856, 1739, 1689, 1610, 1512, 1252, 1173 cm⁻¹; ¹H-NMR (400 MHz, DMSO-d₆, 120°C) δ 0.07 (6H, s), 0.90 (9H, s), 1.16 (3H, s), 1.23 (3H, s), 1.44 (9H, s), 1.45 (3H, s), 2.64 (2H, m), 2.73 (3H, s), 3.11 (1H, d, J = 11.5 Hz), 3.61 (1H, d, J = 11.5 Hz), 3.69 (3H, s), 3.70 (2H, s),3.75 (3H, s), 4.31 (1H, d, J = 3.4 Hz), 4.46 (1H, br-s), 6.85 (2H, d, J = 8.3 Hz), 7.22 (2H, d, J = 8.3 Hz); LSIMS m/z 655 [M+H]⁺, 555, 121; HR-LSIMS m/z 655.3276 [M+H]⁺ (calcd 655.3271 for $C_{32}H_{55}N_2O_6S_2Si$).

N-Me-alcohol 18. A mixture of 17 (304 mg, 0.46 mmol), THF (4.60 ml), MS-4A (600 mg, x 2 wt) and TBAF (1.00 M in THF, 1.16 ml, 2.50 eq.) was stirred at room temperature for 1.5 h. The MS-4A was removed by filtration and the AcOEt filtrate was washed with 5% aq.KHSO₄ and brine, dried over Na₂SO₄, then concentrated under reduced pressure. The residue was dissolved in CH₂Cl₂ (5.00 ml) and MeOH (1.00 ml), and treated with (trimethylsilyl)diazomethane (2.00 M in hexane) dropwise, until generation of N₂ gas ceased. The reaction was quenched with AcOH, and concentrated under reduced pressure. The residue was purified by chromatography (silica 15 g, AcOEt /hexane = 1/5) to give 18 (213 mg, 85%). [α]²⁴_D -73.5 (c 0.40, CHCl₃); IR ν_{max} 3322, 2975, 2835, 1735, 1684, 1610, 1512, 1248, 1171 cm⁻¹; ¹H-NMR (400 MHz, DMSO-d₆, 130°C) δ 1.21 (6H, s), 1.44 (9H, s), 1.45 (3H, s), 2.67 (1H, dd, J = 13.2, 5.4 Hz), 2.73 (1H, m), 2.74 (3H, s), 3.13 (1H, d, J = 11.2 Hz), 3.60 (1H, d, J = 11.2 Hz), 3.69 (2H, s), 3.70 (3H, s), 3.75 (3H, s), 3.84 (1H, d, J = 3.4 Hz), 4.30 (1H, br-s), 6.85 (2H, d, J = 8.5 Hz), 7.22 (2H, d, J = 8.5 Hz); LSIMS m/z 541 [M+H]⁺, 441, 121; HR-LSIMS m/z 541.2403 [M+H]⁺ (calcd 541.2406 for C₂₆H₄₁N₂O₆S₂).

Thiol 20. To an ice-cold solution of **18** (161 mg, 0.30 mmol) in CH_2Cl_2 (6.00 ml) was added freshly prepared Npys-Cl (85.0 mg, 1.50 eq.), and stirred at same temperature for 30 min. The reaction mixture was concentrated under reduced pressure and the residue was chromatographed (silica 10 g, AcOEt /hexane = 2/3 to 3/1) to give an oily product (**19**, 150 mg).

To a solution of 19 (150 mg, ca.0.26 mmol) in acetone (4.00 ml) and water (1.00 ml) was added n-Bu₃P (65.0 μ l, 1.00 eq.), and stirred at room temperature for 20 min. The acetone

was removed in vacuo and the residue was taken up in AcOEt. The AcOEt extract was washed with 10% aq.citric acid and brine, dried over Na₂SO₄, then concentrated under reduced pressure. The residue was purified by chromatography (silica 8.0 g, AcOEt /hexane = 1/3) to afford **20** (83.0 mg, 66% from **18**). [α]²⁵_D -73.2 (c 0.50, CHCl₃); IR ν _{max} 3333, 2974, 2560, 1735, 1690, 1601, 1445, 1366, 1168 cm⁻¹; ¹H-NMR (400 MHz, DMSO- d_6 , 120°C) δ 1.21 (3H, s), 1.22 (3H, s), 1.43 (9H, s), 1.45 (3H, s), 1.80 (1H, t, J = 7.6 Hz), 2.73 (1H, br-d, J = 13.4 Hz), 2.77 (3H, s), 2.81 (1H, m), 3.15 (1H, d, J = 11.5 Hz), 3.60 (1H, d, J = 11.5 Hz), 3.71 (3H, s), 3.86 (1H, d, J = 3.7 Hz), 4.13 (1H, br-s); LSIMS m/z 421 [M+H]⁺, 347, 321, 230; HR-LSIMS m/z 421.1838 [M+H]⁺ (calcd 421.1831 for $C_{18}H_{33}N_2O_5S_2$).

Micacocidin methyl ester 24. To an ice-cold solution of **20** (40.0 mg, 0.10 mmol) in CH_2Cl_2 (2.00 ml) was added TFA (0.40 ml), and stirred at the same temperature for 10 min, then at room temperature for 1 h. The resulting mixture was then concentrated under reduced pressure to afford TFA salt **22** (62.0 mg, quant.) as a pale yellow oil. The product **22** thus obtained was subjected to the next reaction without purification. **22**; IR v_{max} 3308, 2976, 2444, 1739, 1683, 1436, 1203, 1136 cm⁻¹; ¹H-NMR (270 MHz, CDCl₃) δ 1.29 (1H, m), 1.33 (3H, s), 1.48 (3H, s), 1.58 (3H, s), 2.81 (3H, br-s), 2.98 (2H, m), 3.12 (1H, m), 3.24 (1H, d, J = 11.6 Hz), 3.60 (1H, d, J = 11.6 Hz), 3.81 (3H, s), 3.97 (1H, br-s), 7.95 (3H, br-s).

To an ice-cold solution of 23 [1] (31.0 mg, 5.42x10⁻⁵ mol) in THF (1.50 ml) was added LiAlH₄ (2.50 mg, 1.20 eq.), and the mixture was stirred at the same temperature for 30 min. The reaction was quenched with AcOEt, and then the mixture was poured into sat.aq.NH₄Cl and extracted with AcOEt. The AcOEt layer was washed with water and brine, dried over Na₂SO₄, then concentrated under reduced pressure.

Under a nitrogen atmosphere, to a suspension of the residue and AcOK (80.0 mg, 15.0 eq.) in CH₂Cl₂ (1.50 ml) was added a solution of TFA salt 22 (42.0 mg, 1.50 eq.) in CH₂Cl₂ (1.00 ml) dropwise over 30 min, and the mixture was stirred at room temperature for 19 h. The reaction mixture was diluted with AcOEt, washed with 5% aq.KHSO₄ and brine, and dried over Na₂SO₄. Evaporation of the AcOEt phase under reduced pressure gave a pale yellow oil (55.0 mg).

To an ice-cold solution of the oil (55.0 mg) in THF (2.00 ml) was added TBAF (1.00 m in THF, 65.0 μ l, 1.20 eq.), and stirred at room temperature for 15 min. The reaction mixture was diluted with AcOEt and washed with 5% aq.KHSO₄ and brine, dried over Na₂SO₄, then concentrated under reduced pressure. The residue was purified by chromatography (silica 5.0 g, AcOEt /hexane = 2/5) to give micacocidin methyl ester (24) (14.4 mg, 46 % from 23), as a mixture of 4 diastereomers. IR v_{max} 3333, 2957, 2870, 1735, 1582, 1449, 1289, 1207 cm⁻¹; ¹H-NMR (270 MHz, CDCl₃) [major isomer; 9*R*,10*R*] δ 0.90 (3H, t, *J* = 6.9 Hz), 1.31 (3H, s), 1.34 (3H, s), 1.36 (4H, m), 1.52 (3H, s), 1.61 (2H, m), 2.63 (3H, s), 2.92 (1H, dd, *J* = 11.7, 4.5 Hz), 2.95 (2H, m), 3.11 (1H, d, *J* = 11.4 Hz), 3.14 (1H, dd, *J* = 11.4, 7.6 Hz), 3.24 (1H, dd, *J* = 11.7, 6.9 Hz), 3.40 (1H, td, *J* = 6.8, 4.5 Hz), 3.46 (1H, dd, *J* = 11.4, 8.7 Hz), 3.51 (1H, d,

J = 6.6 Hz), 3.62 (1H, d, J = 11.4 Hz), 3.78 (3H, s), 4.19 (1H, d, J = 9.2 Hz), 4.77 (1H, ddd, J = 9.2, 8.7, 7.6 Hz), 6.70 (1H, dd, J = 7.6, 1.3 Hz), 6.85 (1H, dd, J = 8.2, 1.3 Hz), 7.21 (1H, dd, J = 8.2, 7.6 Hz).

Micacocidin (1). To an ice-cold solution of micacocidin methyl ester 24 (a mixture of 4 diastereomers, 12.0 mg, 2.06x10⁻⁵ mol) in THF (1.00 ml) and water (0.25 ml) was added LiOH-H₂O (1.80 mg, 2.10 eq.), and stirred at room temperature for 30 min. The reaction mixture was diluted with AcOEt and washed with 5% aq.KHSO₄ and brine, dried over Na₂SO₄, then concentrated under reduced pressure to give crude micacocidin as a mixture of 4 diastereomers.

To a solution of the residue in MeOH (1.00 ml) and water (0.25 ml) was added ZnCl₂ (42.0 mg, 15.0 eq.), and stirred at room temperature overnight. The mixture was diluted with AcOEt and washed with 5% aq.KHSO₄ to release the zinc ion. The organic layer was washed with brine, dried over Na₂SO₄, then concentrated under reduced pressure to give crude micacocidin (8.50 mg, ca.3:1 mixture of 2 diastereomers of C-9 configuration).

The crude micacocidin (6.0 mg) was purified by HPLC [ODS HG-5 (50 x 250 mm), 75 % MeOH + 1 mM phosphate buffer (pH 7), 7.5 ml/min, det. UV 254 nm] to yield pure micacocidin (3.0 mg, Rt; 20.3 min) and C-9 isomer (1.2 mg, Rt; 18.5 min). 1; $[\alpha]^{22}_{D}$ -65.3 (c 0.93, MeOH); IR v_{max} 3064, 2924, 2857, 1729, 1581, 1464, 1290, 1211 cm⁻¹; ¹H-NMR (270 MHz, CDCl₃) δ 0.90 (3H, t, J = 6.9 Hz), 1.31 (3H, s), 1.32 (3H, s), 1.36 (4H, m), 1.58 (3H, s), 1.61 (2H, m), 2.63 (3H, s), 2.94 (3H, m), 3.16 (1H, dd, J = 11.4, 7.6 Hz), 3.17 (1H, d, J = 11.5 Hz), 3.26 (1H, dd, J = 11.2, 7.2 Hz), 3.34 (1H, td, J = 6.9, 4.0 Hz), 3.47 (1H, dd, J = 11.4, 8.7 Hz), 3.61 (1H, d, J = 6.8 Hz), 3.65 (1H, d, J = 11.5 Hz), 4.22 (1H, d, J = 9.1 Hz), 4.76 (1H, td, J = 8.9, 7.6 Hz), 6.64 (1H, br-s), 6.71 (1H, dd, J = 7.6, 1.2 Hz), 6.86 (1H, dd, J = 8.2, 1.2 Hz), 7.21 (1H, t, J = 7.8 Hz); HR-LSIMS m/z 566.2183 [M+H]⁺ (calcd 566.2179 for $C_{27}H_{39}N_3O_4S_3$).

Acknowledgments

We are very grateful to Professor Isao Kitagawa (Kinki University, Osaka) for invaluable discussion and helpful support. We also thank Mr. Shinobu Kobayashi for his advice regarding HPLC purification of micacocodin.

References

- [1] Ino, A.; Murabayashi, A. Tetrahedron, preceding paper.
- [2] a) Shionogi & Co., Ltd. Patent, WO 96/04262. b) Kobayashi, S.; Hidaka, S.; Kawamura, Y.; Ozaki, M.; Hayase, Y. J. Antibiotics 1998, 51, 323-327. c) Kobayashi, S.; Nakai, H.; Ikenishi, Y.; Sun, Y.; Ozaki, M.; Hayase, Y.; Takeda, R. ibid. 1998, 51, 328-332.
- [3] Ino, A.; Hasegawa, Y.; Murabayashi, A. Tetrahedron Lett. 1998, 39, 3509-3512.

- [4] Ino, A.; Kobayashi, S.; Hidaka, S.; Kawamura, Y.; Ozaki, M.; Hayase, Y.; Takeda, R.; Murabayashi, A. The 38th Symposium on the Chemistry of Natural Products, Symposium Papers 1996, 121-126.
- [5] Kemp, D.S.; Carey, R.I. J. Org. Chem. 1989, 54, 3640-3646.
- [6] Onoda, T.; Shirai, R.; Koiso, Y.; Iwasaki, S. Tetrahedron Lett. 1995, 36, 5765-5768.
- [7] a) Mulqueen, G.C.; Pattenden, G.; Whiting, D.A. *Tetrahedron* 1993, 49, 5359-5364. b) Pattenden, G.; Thom, S.M.; Jones, M.F. *ibid.* 1993, 49, 2131-2138.
- [8] Ishizuka, T.; Kunieda, T. Tetrahedron Lett. 1987, 28, 4185-4188.
- [9] a) Matsueda, R.; Higashida, S.; Ridge, R.J.; Matsueda, G.R. Chem. Lett. 1982, 921-924. b) Pugh, K.C.; Gera, L.; Stewart, J.M. Int. J. Peptide Protein Res. 1993, 42, 159-164.